

A chemotactic model of bacterial aggregation with nonlinear cross diffusion

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Joint work with:

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① Introduction

② Modelling

③ Asymptotics for the envelope front speed

④ Numerical simulations

Bacterial aggregation patterns

- Bacterial colonies *in vitro* exhibit morphological aggregation patterns
- Hostile environmental conditions: low nutrient level, hard agar
- Adaptive survival strategies lead to complex spatio-temporal patterns.
- Complex patterns; complex self-organization: micro-level (cell-cell), macro-level (colony), chemical signalling, gene exchange.
- Example: *Bacillus subtilis*. Gram + bacteria, resistant.

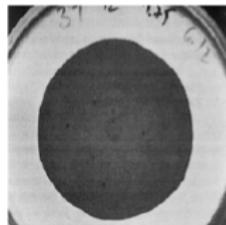
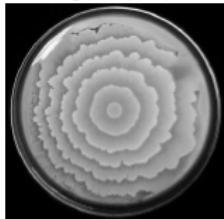
Observations

- Low nutrient, hard agar: Diffusion limited aggregation (DLA). Fractal patterns (Matsuyama and Matsushita (1993); Ben-Jacob (1994)). **(A)**
- Semi-solid agar, low nutrient: Dense branch morphology (DBM). Smooth colony envelope (Ohgiwari et al. (1992)). **(E)**
- Higher nutrient, soft agar: homogeneous colony, smooth boundary envelope. **(D)**
- Hard agar, high nutrient: envelope with fractal boundary. **(B)**
- Rings: transition from **(B)** to **(D)**.

C_n - concentration of nutrient; C_a - agar concentration
(softness $1/C_a$)

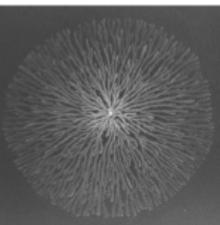
Morphological diagram

Rings (C)

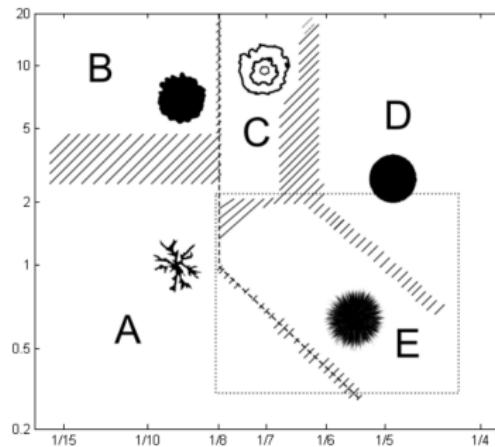


Disk (D)

Fractal (A)



DBM (E)

 C_n vs. $1/C_a$

Deterministic reaction-diffusion models

- Continuous time evolution systems of PDEs
- Reaction-diffusion modelling (Murray)
- Non-equilibrium patterns: colonies in invaded and non-invading regions
- Diffusive growth: the boundary of the colony is unstable
- Traveling wave approximations, speed of colony envelope.
- Toy model (paradigm): Fisher-KPP one -d reaction-diffusion equations (propagating front)
- Other patterns: Turing diffusion-driven linear instability

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Kawasaki's model

Kawasaki et al. *J. Theor. Biol.* **188** (1997): Non-linear cross-diffusion model

$$\begin{aligned} n_t &= D_n \Delta n - f(n, b) \\ b_t &= \nabla \cdot (D_b \nabla b) + \theta f(n, b) \end{aligned} \tag{K}$$

n - nutrient and b - bacterial concentrations,
 $(x, y) \in \Omega \subset \mathbb{R}^2$, $t > 0$. Diffusion: $D_n > 0$ constant.

Consumption rate: $f(n, b)$, $\theta > 0$ -conversion rate factor.

No-flux boundary conditions.

Nonlinear cross-diffusion

$$D_b = \sigma n b$$

$\sigma = \sigma_0(1 + \Delta)$. $\sigma_0 \sim 1/C_a$. Δ - random parameter

Features:

- Ohgiwari et al., (1992): low and intermediate C_a , growth by mov. cells. Low agar. Boundary front: cells are active on the bdry. *only*.
- Kawasaki: immotility when either n or b are low.
- Complex dense morphology. Rich mathematical structure (Murray; Satnoianu, Maini and Sánchez-Garduño (2001); Sherratt (2010)).

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Chemotaxis

E. Ben-Jacob's group (Univ. of Tel-Aviv):

- Ben-Jacob et al. *Nature* **368** (1994): Experimental evidence of chemotaxis towards amino acids (nutrients)
- Golding et al. *Phys. A* **260** (1997): Role of chemotaxis in the formation of patterns; theoretical and experimental arguments
- Ben-Jacob et al. *Adv. Phys.* **49** (2000): Identified three types of chemotactic internal signalling. Propose, based on experiments, a relation bet. diffusion and chemotaxis (certain nutrient regimes)

Chemotactic responses:

- (Repulsive - Long range) By starving bacteria in the center.
- (Attractive - Short range) Bacteria in the front ask for help to metabolize waste.
- **Nutrient chemotaxis:** Dominant signal. Attractive, short range.

Chemotactic flux

$$J_c = \xi(b)\chi(n)\nabla n.$$

$\chi = \chi(n) > 0$ - chemotactic sensitivity function

$\xi = \xi(b)$ - bacterial response function to nutrient gradient

- $\xi > 0$ - attractive
- $\xi < 0$ - repulsive

Receptor's law

Lapidus-Schiller (1976):

$$\chi(n) = \frac{\chi_0 K_d}{(K_d + n)^2}$$

- $\chi_0 > 0$: constant (strength of the chemotaxis)
- $K_d > 0$: receptor-ligand binding dissociation constant.
Units of n ; concentration 1/2 of receptor to be
occupied. Unique value under fixed conditions.

First approximation: n small, $\chi(n) \sim \chi_0 / K_d$.

Ben-Jacob's conjecture

In semi-solid agar, low colony density:

$$|\xi(b)| \propto bD_b.$$

Examples:

- Keller-Segel (1971): $D_b > 0$ constant \Rightarrow
 $\xi(b) = \pm bD_b.$
- Kitsunezaki (1997): $D_b = D_0 b^m$, $m > 0$, $D_0 > 0$ const.
 $\Rightarrow \xi(b) = bD_b = D_0 b^{m+1}$

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Chemotactic flux

$$D_b = \sigma n b$$

$$J_c = \sigma n b^2 \chi(n) \nabla n,$$

Positive (attractive) bacterial response function. χ is either constant or the Lapidus-Schiller receptor law.

Chemotactic model with nonlinear cross diffusion

With receptor's law:

$$n_t = D_n \Delta n - \kappa n b,$$

$$b_t = \nabla \cdot (\sigma n b \nabla b) + \theta \kappa n b - \nabla \cdot \left(\sigma n b^2 \frac{\chi_0 K_d}{(K_d + n)^2} \nabla n \right),$$

$$(x, y) \in \Omega \subset \mathbb{R}^2, t \geq 0. \quad \sigma = \sigma_0(1 + \Delta).$$

- No-flux boundary conditions

$$\nabla b \cdot \mathbf{v} = 0, \quad \nabla n \cdot \mathbf{v} = 0, \quad (x, y) \in \partial\Omega$$

- Initial conditions

$$n(x, y, 0) = \hat{n}_0, \quad b(x, y, 0) = \hat{b}_0(x, y)$$

$\hat{n}_0 > 0$ constant, \hat{b}_0 given.

Non-dimensionalization

$$\tilde{x} = \left(\frac{\theta K_d \kappa}{D_n} \right)^{1/2} x, \quad \tilde{y} = \left(\frac{\theta K_d \kappa}{D_n} \right)^{1/2} y, \quad \tilde{t} = (\theta K_d \kappa) t,$$

$$\tilde{n} = \frac{n}{K_d}, \quad \tilde{b} = \frac{b}{\theta K_d}, \quad \tilde{\sigma} = \left(\frac{\theta K_d^2}{D_n} \right) \sigma.$$

$$n(x, y, 0) = \hat{n}_0 / K_d \equiv n_0,$$

$$b(x, y, 0) = \hat{b}_0(x, y, 0) / (\theta K_d) \equiv b_0(x, y, 0),$$

$$n_t = \Delta n - nb,$$

$$b_t = \nabla \cdot (\sigma nb \nabla b) + nb - \chi_0 \nabla \cdot \left(\frac{\sigma nb^2}{(1+n)^2} \nabla n \right). \quad (C)$$

Initial cond. $n(x, y, 0) = n_0$, $b(x, y, 0) = b_0(x, y)$. No flux b.c.

Free parameters:

- $\sigma_0 > 0$ - hardness of the agar medium (large σ_0 , soft agar)
- $n_0 > 0$ - relative initial nutrient
- $\chi_0 \geq 0$ - intensity of the chemotaxis

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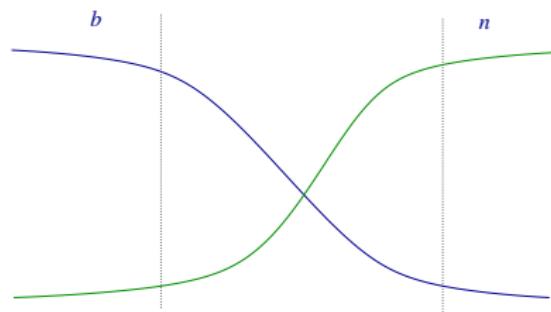
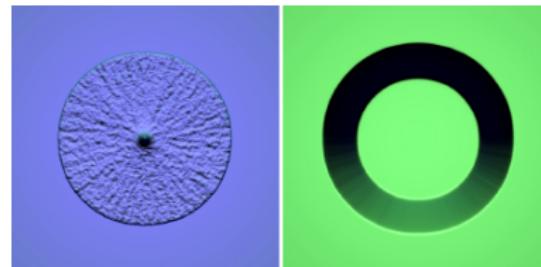
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Approximation:

- Conservation law: $b + n \approx C$ constant.
- $n = n_0 - b$ leads to a logistic equation, absence of diffusion and chemotaxis
- $\chi(n) \approx \chi_0$
- $\sigma = \sigma_0$



Front approximation: $n \approx n_0 - b$

Approximate scalar equation near front:

$$b_t = \sigma_0 \nabla \cdot (\tilde{D}(b, \chi_0) \nabla b) + g(b)$$

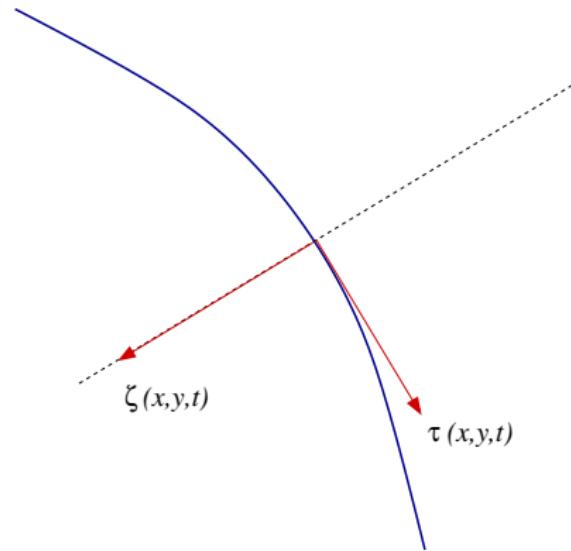
$$\tilde{D}(b, \chi_0) = n_0 b \left(1 - \frac{b}{n_0} \right) (1 + \chi_0 b)$$

$$g(b) = n_0 b \left(1 - \frac{b}{n_0} \right)$$

- Scalar equation for b with Fisher-type reaction term $g(b)$
- Effective non-linear diffusion coefficient $\tilde{D}(b, \chi_0)$

Geometric front propagation

- Local curvilinear coordinates: normal $\zeta(x, y, t)$, tangent $\tau(x, y, t)$
- $b(x, y, t) \approx b(\zeta(x, y, t))$ motion in the normal direction



Front equation

$$-cb' = \sigma_0(\tilde{D}(b)b')' + b(n_0 - b)$$

$$c = -\zeta_t + \sigma_0 \tilde{D}(b) \Delta \zeta$$

Theoretical results

- Malaguti and Marcelli, *J. Diff Equations* **195** (2003)

Fisher-KPP equation with doubly degenerate diffusion:

$$u_t = (D(u)u_x)_x + g(u)$$

$$0 < u < 1, D \in C^1([0, 1]), g \in C^1([0, 1])$$

$$D(0) = D(1) = 0, \quad D(u) > 0 \text{ for all } 0 < u < 1,$$

$$g(0) = g(1) = 0, \quad g(u) > 0 \text{ for all } 0 < u < 1,$$

Existence of TWS provided the speed c satisfies:

$$0 < c_* \leq c$$

Threshold velocity c_* of a sharp front:

$$0 < c_* \leq 2 \sqrt{\sup_{s \in (0,1)} \frac{D(s)g(s)}{s}}.$$

Front equation:

$$b_t = (\tilde{D}(b, \chi_0)b_x)_x + g(b),$$

$b \in [0, n_0]$, $\tilde{D}(b, \chi_0)$, $g(b)$ chemotactic dependent $\chi_0 \geq 0$.
 Existence of TWS linked to the solution to the b.v. problem:

$$\frac{dz}{db} = -c - \frac{\tilde{D}(b, \chi_0)g(b)}{z},$$

$$z(0^+) = z(n_0^-) = 0.$$

$A_{\chi_0} = \{c > 0 : \text{b.v. problem has a non-positive solution}\}.$

$$c_* = \inf A_{\chi_0}$$

Lemma

For any fixed value of $n_0 > 0$ and for all values of $\chi_0 \geq 0$ there hold,

(i) $\bar{c}(\chi_0) \geq \bar{c}(0)$, and

(ii) $c_*(\chi_0) \geq c_*(0)$

where

$$\bar{c}(n_0, \chi_0) = 2n_0 \sqrt{\sigma_0} \sqrt{\max_{b \in [0, n_0]} b \left(1 - \frac{b}{n_0}\right)^2 (1 + \chi_0 b)}.$$

Let

$$\psi(b) = b(1 - b/n_0)^2(1 + \chi_0 b) \geq 0$$

for $b \in [0, n_0]$, each $\chi_0 \geq 0$. Global max:

$$\psi_{\max} = \psi(b_*),$$

$$b_* = \frac{1}{2} \left(\frac{n_0}{2} - \frac{3}{4\chi_0} \right) + \frac{1}{2} \sqrt{\left(\frac{n_0}{2} - \frac{3}{4\chi_0} \right)^2 + \frac{n_0}{\chi_0}} \in (0, n_0).$$

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$$\bar{c}(n_0, \chi_0) = 2n_0 \sqrt{\sigma_0} \sqrt{\psi(b_*)}.$$

No chemotaxis $\chi_0 = 0$: Newman (1980), $\psi_{\max} = 4n_0/27$

$$\bar{c}(n_0, 0) = 2n_0 \sqrt{\sigma_0} \sqrt{\frac{4n_0}{27}} = \frac{4}{3\sqrt{3}} n_0^{3/2} \sigma^{1/2}$$

Bounds for the speed of the sharp front. In the approximation $\kappa = -\Delta \zeta > 0$ (curvature), so that

$$s = -\zeta_t \geq c_*(\chi_0) + \sigma_0 \tilde{D}(b) \kappa > 0.$$

Approximate interface equation of motion with chemotactic contribution:

$$s = -\zeta_t \geq c_*(\chi_0) + \sigma_0 \tilde{D}(b) \kappa \geq c_*(0) + \sigma_0 \tilde{D}(b) \kappa \geq c_*(0)$$

Numerical approximation of the velocity

Consider the one-d version:

$$n_t = (n_x)_x - nb,$$

$$b_t = (\sigma_0 n b b_x - \chi_0 \sigma_0 n b^2 n_x)_x + nb.$$

Simulations with

$$n(x, 0) = n_0, \quad b(x, 0) = b_M e^{-x^2/6.25},$$

where $b_M = 0.71$. Underlies approximate traveling wave profile solutions for b ; approximate speed can be computed from the numerical solution.

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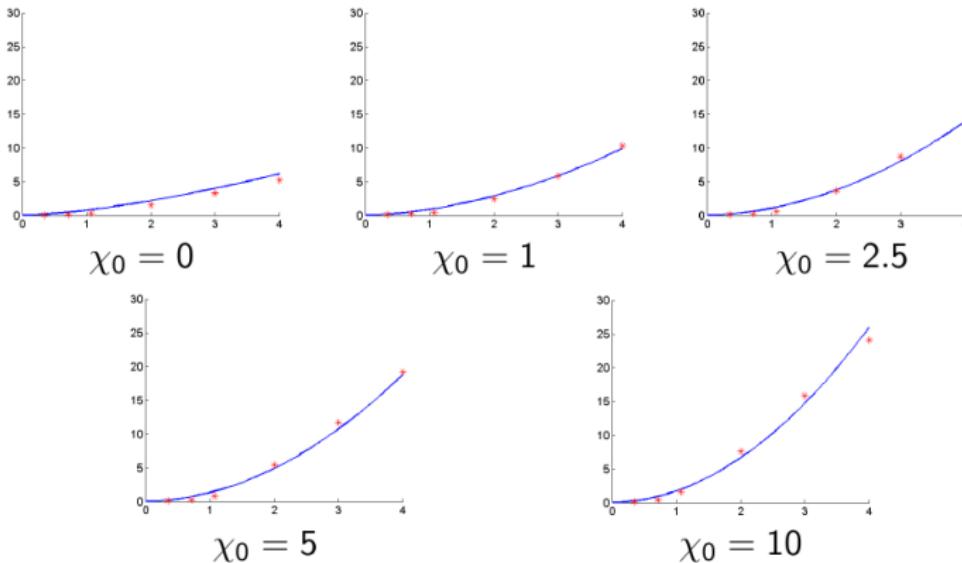


Figure: Propagation speeds vs. n_0 : * = numerical estimation;
solid line = $2n_0\sigma_0^{1/2}\psi(b_*)^{1/2}$, $\sigma_0 = 1$

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Scheme

- Square domain $[0, 1] \times [0, 1]$. Grid of 2048×2048
- Finite difference, 2nd. order Runge-Kutta scheme
- Very small time steps to avoid instabilities (stiffness)
- Compensated by parallel high performance computations with 100s of processors in Graphic Processing Units (GPUs)
- Millions of steps in a few hours.
- NVIDIA Tesla[©] C2070 graphics card with 448 CUDA cores



Parameter values:

Description	Symbol	Values
Initial condition for the nutrient	n_0	1.07, 0.71
Agar softness	σ_0	1.0, 4.0
Chemotactic signal strength	χ_0	0, 2.5, 5.0, 7.5, 10.0

Initial condition (Kawasaki):

$$b_0(x, y, 0) = b_M e^{-(x^2 + y^2)/6.25}, \quad b_M = 0.71.$$

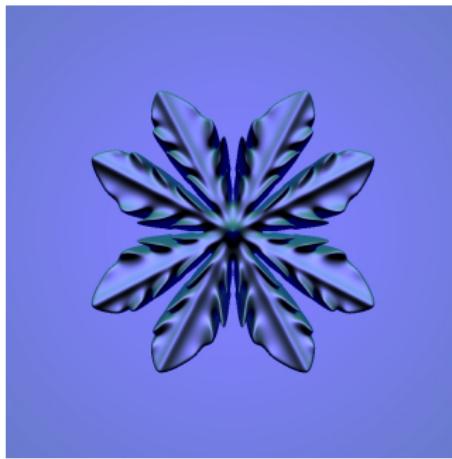


Figure: Simulations with $n_0 = 0.71$, $\chi_0 = 5$, no random fluctuations $\Delta = 0$ ($\sigma = \sigma_0 = 1$); grid of 2048×2048 . High degree of symmetry due to the implicit anisotropy of the grid.

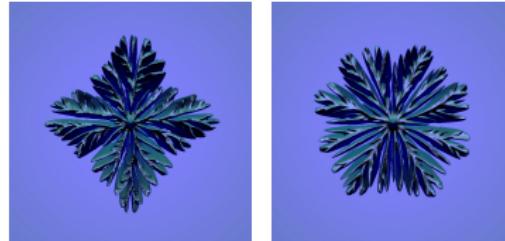
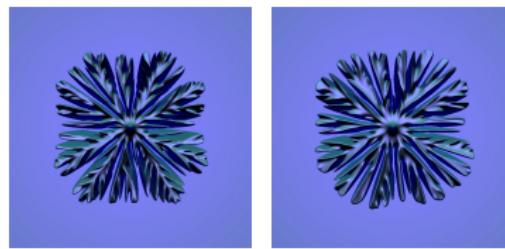
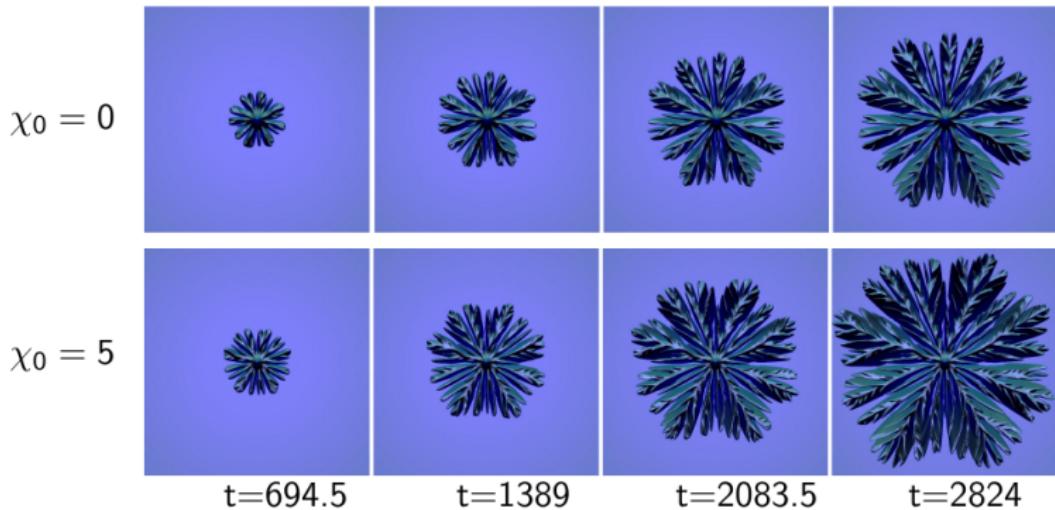
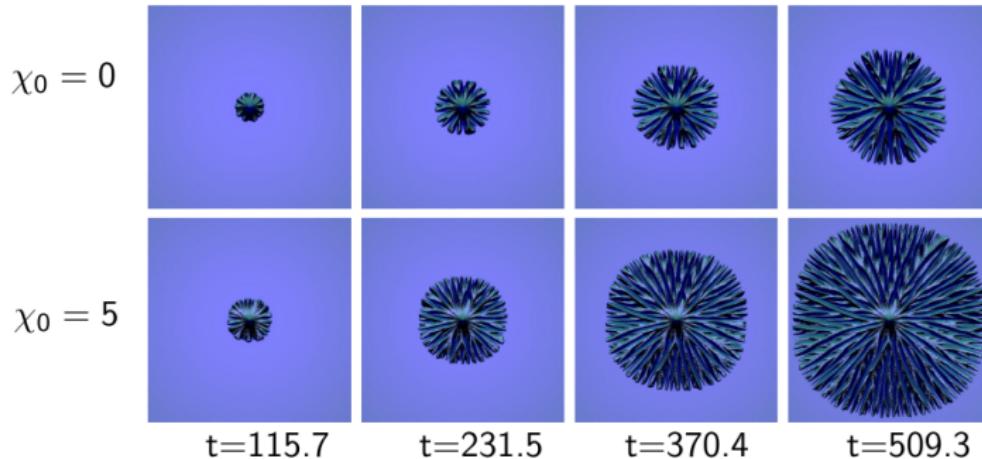
(a) 1024×1024 (b) 2048×2048 (c) 4096×4096 (d) 8192×8192

Figure: Simulations with $\sigma_0 = 1, n_0 = 0.71$ and $\chi_0 = 5, \Delta \neq 0$.

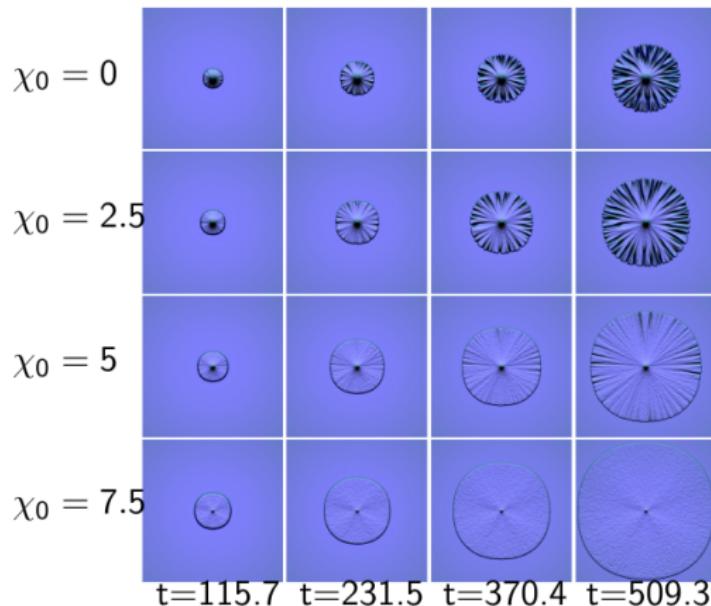
Semi-solid agar: $\sigma_0 = 1$; low-nutrient $n_0 = 0.71$.



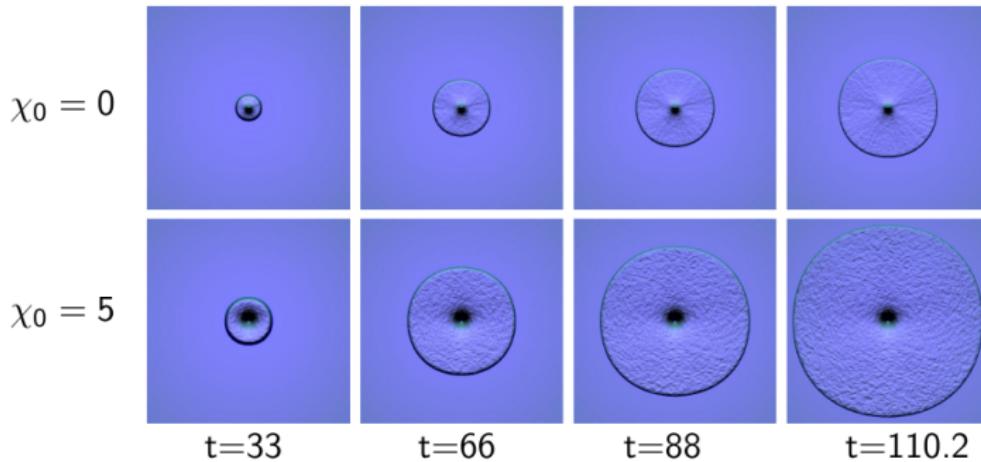
Semi-solid agar: $\sigma_0 = 1$; medium-nutrient $n_0 = 1.07$.



Soft agar: $\sigma_0 = 4$; low-nutrient $n_0 = 0.71$.



Soft agar: $\sigma_0 = 4$; med-nutrient $n_0 = 0.71$.



Discussion

- Incorporation of a suitable chemotactic term to Kawasaki's nonlinear cross diffusion model
- Compatible with low-nutrient regime experimental observations (Ben-Jacob)
- Asymptotics show that the speed of the envelope front increases with chemotaxis
- Numerical estimation (one-d simulation) approximates well the asymptotic speed calculation
- High resolution numerical simulations confirm enhancement of the speed
- In the low-nutrient, soft-agar regime: change in morphology, patterns become smoother (less branches) in the presence of chemotaxis

Instability vs. front speed

Transition (**E**) to (**D**). Effect of chemotaxis supresses the onset of instability (formation of branches). Morphology gets homogeneized. Still, speed of front increases.

- Arouh, Levine, *Phys. Rev E*. **62** (2000): for Kessler-Levine RD equations, add nutrient chemotaxis. Reflected in the decrease of the “growth rate” of the perturbation of the envelope: supress instability. Calculation can be extrapolated to other models.

“...Why does chemotaxis suppress the onset of the instability, especially inasmuch as a naive analysis might lead one to argue that chemotaxis causes outwardly protruding parts of the colony to move faster as they feel higher gradients?”

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Reference:

- Leyva, Malaga, P, *Phys. A* **392** (2013), 5644-5662.

Thank you!